

The Effect of Fenugreek (*Trigonella foenum-graecum*) Seed and 17- β Estradiol on Serum Apelin, Glucose, Lipids, and Insulin in Ovariectomized Rats

Mahmood Abedinzade¹; Sima Nasri²; Masome Jamal Omid²; Bizhan Porramezan²; Korosh Khanaki^{3,*}

¹Medical Biotechnology Research Center, Department of Medical Physiology, Faculty of Paramedicine, Guilan University of Medical Sciences, Rasht, IR Iran

²Department of Biology, Payame Noor University, Tehran, IR Iran

³Medical Biotechnology Research Center, Department of Clinical Biochemistry, Faculty of Paramedicine, Guilan University of Medical Sciences, Rasht, IR Iran

*Corresponding author: Korosh Khanaki, Medical Biotechnology Research Center, Department of Clinical Biochemistry, Faculty of Paramedicine, Guilan University of Medical Sciences, Rasht, IR Iran. Tel: +98-1342536767, Fax: +98-1342537070, E-mail: khanaki_korosh_bio@yahoo.com

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Background: Menopause, a natural phenomenon, is defined by the fall of ovarian hormones mainly estrogens causing major problems such as insulin resistance. Fenugreek (*Trigonella foenum-graecum*) is known to have some useful properties such as insulin sensitizing effect. Apelin is an adipokine, which has several roles such as regulation of insulin secretion.

Objectives: The objective of the present study was to evaluate the effect of fenugreek seed and 17- β estradiol on serum Apelin along with glucose, lipids and insulin in ovariectomized rats.

Materials and Methods: Forty-nine adult female Wistar rats were randomly divided to seven groups: normal control, ovariectomized control, ovariectomized treated with ethanolic and hexanic extract of fenugreek seed (50 and 150 mg/kg/daily for each), and ovariectomized treated with 17- β estradiol (10 μ g/kg/daily) for 42 days. Serum Apelin, glucose, lipids and insulin were measured.

Results: Serum Apelin, glucose, lipids and insulin significantly increased in ovariectomized controls in comparison with normal controls ($P < 0.05$). Serum glucose, lipids and insulin in ovariectomized rats treated with fenugreek seed extract and 17- β estradiol were remarkably lower than ovariectomized controls ($P < 0.05$). Furthermore, 17- β estradiol caused a significant decrease ($P < 0.05$) in serum Apelin in ovariectomized rats.

Conclusions: It appears that fenugreek seed might be effective against hyperglycemia, hyperlipidemia and insulin resistance in ovariectomized rats without impact on serum Apelin. Furthermore, 17- β estradiol could have similar effects along with possible inhibitory effects on serum Apelin. The complicated role of Apelin in menopause needs to be further explored.

Keywords: Apelin-13; *Trigonella Foenum-Graecum*; 17 Beta-Estradiol; Ovariectomized; Insulin; Lipids

1. Background

Menopause is a natural event characterized by follicular atresia and fall in the secretion of ovarian hormones especially estrogens (1). This condition leads to physiological and biochemical changes in affected females such as increased blood pressure, insulin resistance, and dyslipidemia (2). These changes are associated with increased risk of metabolic syndrome and some chronic diseases such as cardiovascular diseases (3).

Menopause usually occurs in middle-aged women; between 45 and 55 years (4). According to a survey, the number of perimenopausal or postmenopausal females would be about 1.2 billion by the year 2030 (5). Since menopause mostly affects females for about a third of their lives, providing a new effective approach for the prevention of menopause complications or decreasing the risk of associated diseases could be helpful in enhancing health and quality of life among females.

Apelin is an adipokine with a number of active molecular forms, which is expressed in different tissues such as adipocytes (6). Apelin has several important roles such as regulation of insulin secretion through autocrine/paracrine stimulation (7), and inhibition of lipolysis in adipocytes (8). The role of Apelin in lowering blood glucose and insulin sensitivity has been suggested by previous studies (9). Assessment of the possible interaction between Apelin signaling, lipid metabolism, and beta cell function could be worthy of investigation (10). Fenugreek (*Trigonella foenum-graecum*), as an annual medicinal plant, is a member of the Fabaceae family. Fenugreek seed has been used to promote health and as a choice for the management of various human diseases such as diabetes, dyslipidemia and obesity (11, 12). The insulin sensitizing action of fenugreek has been proposed by earlier studies (13).

2. Objectives

For the first time, the present study aimed to evaluate the effect of fenugreek seed and 17- β estradiol on the serum level of Apelin along with glucose, lipids and insulin in ovariectomized (OVX) rats as a model of menopause.

3. Materials and Methods

3.1. Animals

Forty-nine adult female Wistar rats weighing 190 - 220 g, obtained from the Institute of Pasteur, were used. Ethical clearance from this experimental study was approved by the ethical committee at Guilan university of medical sciences and the experiments were done in agreement with internationally accepted principles for laboratory animal use and cares as found in US guidelines (NIH publication #85-23, revised in 1985). Animals were housed in a room maintained at 21 to 23°C with a 12 hours light/dark cycle. All rats had free access to food and water.

3.2. Study Design

After an adaptation period of two weeks, all rats were randomly divided to seven groups (seven rats in each group) as follows:

1) Normal control; receiving daily saline.

The remaining six groups were allocated to undergo bilateral ovariectomy as follows (14):

2) OVX control; receiving daily saline.

3) OVX treated daily with 50 mg/kg of ethanolic extract of fenugreek seed (EEFS).

4) OVX treated daily with 150 mg/kg of EEFS.

5) OVX treated daily with 50 mg/kg of hexanic extract of fenugreek seed (HEFS).

6) OVX treated daily with 150 mg/kg of HEFS.

7) OVX treated daily with 10 μ g/kg of estradiol.

Treatment began one day after ovariectomy by Intraperitoneal (IP) injection and all rats were maintained for 42 days on their respective treatments. The weight and fasting blood sugar (FBS) of all rats were measured at the beginning of the research and on the 42nd day of treatment. Blood glucose was measured by a glucometer (Accu chek, Roche, Germany). After 42 days, fasting whole blood samples were obtained from the vein of the tail. The serum of blood samples was separated immediately. The resulting sera were stored at -20°C. The serum levels of Apelin-13 and insulin were determined by using enzyme-linked immunosorbent assay (ELISA) kits (Shanghai Crystal Day Biotech Co., China). The serum levels of cholesterol, triglyceride (TG), low-density Lipoprotein (LDL) and high-density lipoprotein (HDL) were determined by using commercial kits (Pars Azmoon, Iran).

3.3. Plant Material and Extraction

The seed of fenugreek was supplied by a grocery store in Rasht (Guilan province) during years 2013 and 2014, and the species was confirmed by the herbarium department of the agricultural research center at Guilan university. The voucher specimen 11411 was deposited in the Herbarium of the department of botany, faculty of agriculture, Guilan university. Preparation of the ethanolic and hexanic extracts was performed as previously described (11).

3.4. Chemicals

The 17- β estradiol was obtained from Sigma-Aldrich (GmbH, Germany). Ethanol and hexane were purchased from Merck (Germany).

3.5. Statistical Analysis

Data are presented as means \pm standard error of the Mean (SEM) and inter-group comparisons were made using the one-way analysis of variance (ANOVA) followed by post hoc Tukey's test. P values of < 0.05 were considered statistically significant. Analysis was performed using the SPSS software version 16.

4. Results

The average body weight of OVX control remarkably increased in comparison to normal control rats ($P < 0.05$). Treatment of the OVX rats with HEFS, EEFS and estradiol resulted in a decrease in body weight (Table 1).

The serum level of cholesterol, TG and LDL remarkably increased while serum HDL significantly decreased in OVX control when compared to normal control rats ($P < 0.05$). However, HEFS, EEFS, and estradiol caused a remarkable decrease in serum level of cholesterol and TG in OVX rats ($P < 0.05$). Following 42 days of treatment with HEFS at a dose of 150 mg/kg/day, EEFS at a dose of 50 mg/kg/day and estradiol, the serum LDL was reduced. Only 17- β estradiol caused remarkable increase in the serum level of HDL (Table 1).

The serum glucose and insulin significantly increased in OVX controls in comparison with normal control rats ($P < 0.05$) (Table 2).

The HEFS, EEFS, and estradiol rats showed a remarkable decrease in the serum levels of glucose and insulin when compared to OVX control rats ($P < 0.05$) (Table 2).

The serum level of Apelin was significantly higher in OVX control rats compared with normal control rats ($P < 0.05$). There were no significant differences in the level of serum Apelin between OVX rats treated with HEFS and EEFS as compared with OVX control rats. The 17- β estradiol caused a significant decrease ($P < 0.05$) in serum Apelin in ovariectomized rats ($P < 0.05$) (Table 2).

Table 1. Effect of Fenugreek Seed and 17- β Estradiol on Body Weight, Serum Cholesterol, Low-Density Lipoprotein, High Density Lipoprotein, and Triglyceride After Six Weeks in Normal Control and Ovariectomized Rats ^{a,b}

Groups	Body weight, g	Cholesterol, mg/dl	LDL, mg/dl	HDL, mg/dl	Triglyceride, mg/dl
Normal control	233 \pm 2 ^c	85 \pm 2.6	51.7 \pm 4.3	24 \pm 1.92	60.1 \pm 2.4
OVX control	247 \pm 3 ^d	122 \pm 9 ^c	83 \pm 5.1 ^d	12 \pm 2.64 ^d	115 \pm 1.1 ^d
OVX + HEFS (50 mg/kg)	235 \pm 3 ^c	80 \pm 3 ^c	75.1 \pm 4 ^d	18 \pm 1	84.1 \pm 3.9 ^{c,d}
OVX + HEFS (150 mg/kg)	233 \pm 2 ^c	86 \pm 4 ^c	60.2 \pm 1.6 ^c	20 \pm 2.31	71.5 \pm 2.9 ^c
OVX + EEFS (50 mg/kg)	229 \pm 3 ^c	93 \pm 4 ^c	74.4 \pm 3.1 ^c	14 \pm 1.31	88.2 \pm 4.2 ^{c,d}
OVX + EEFS (150 mg/kg)	231 \pm 2 ^c	73 \pm 5 ^c	69.8 \pm 3.5 ^d	16 \pm 2.41	79 \pm 3.9 ^{c,d}
OVX + estradiol (10 μ g/kg)	231 \pm 1 ^c	100 \pm 3 ^{c,d}	54.2 \pm 5.1 ^c	29 \pm 1.81 ^c	73.5 \pm 4.2 ^c

^a Values are given as mean \pm SEM.

^b Abbreviations: EEFS: ethanolic extract of fenugreek seed, HDL: high-density lipoprotein, HEFS: hexanic extract of fenugreek seed, LDL: low-density lipoprotein, OVX: ovariectomized.

^c P < 0.05 by comparison with ovariectomized control rats.

^d P < 0.05 by comparison with normal control rats.

Table 2. Effect of Fenugreek Seed and 17- β Estradiol on Serum Glucose, Insulin and Apelin After Six Weeks in Normal Control and Ovariectomized Rats ^{a,b}

Groups	Glucose, mg/dl	Insulin, U/dl	Apelin, μ g/dl
Normal control	97 \pm 2.04	489 \pm 1.92	75.1 \pm 1.5
OVX control	158 \pm 1.95 ^c	700 \pm 1.64 ^c	88.8 \pm 1.9 ^c
OVX + HEFS (50 mg/kg)	132 \pm 2.18 ^{c,d}	627 \pm 1.21 ^{c,d}	86.2 \pm 3.5
OVX + HEFS (150 mg/kg)	128 \pm 3.18 ^{c,d}	600 \pm 1.81 ^{c,d}	80 \pm 1.9
OVX + EEFS (50 mg/kg)	135 \pm 3.38 ^{c,d}	649 \pm 1.01 ^{c,d}	80.7 \pm 2.4
OVX + EEFS (150 mg/kg)	129 \pm 2.61 ^{c,d}	616 \pm 0.81 ^{c,d}	81.4 \pm 4
OVX + estradiol (10 μ g/kg)	134 \pm 6.18 ^d	546 \pm 2.81 ^{c,d}	72.1 \pm 0.9 ^d

^a Values are given as mean \pm SEM.

^b Abbreviations: EEFS: ethanolic extract of fenugreek seed, HEFS: hexanic extract of fenugreek seed, OVX: ovariectomized.

^c P < 0.05 by comparison with normal control rats.

^d P < 0.05 by comparison with ovariectomized control rats.

5. Discussion

The present study was undertaken to find out the effect of fenugreek seed extract and 17- β estradiol on serum Apelin together with some of menopause related parameters such as serum lipids, glucose, and insulin level. Mean level of serum cholesterol, TG, and LDL in OVX control were significantly higher than those of normal control rats, whereas there was a statistically significant decrease of HDL in OVX control in comparison to normal control rats. These findings were similar to that found by the study of Goss et al. (15) in which ovariectomy caused a considerable increase in serum levels of cholesterol and LDL in comparison to normal rats. In another study (16), OVX rats showed no significant changes in TG, cholesterol and HDL levels compared with normal rats. Meno-

pausal statement due to reduced estrogen secretion is associated with hyperlipidemia, which is defined by increased level of cholesterol, TG, LDL, and decreased level of HDL (17). It has been demonstrated that estrogen has a physiological role in lowering cholesterol through up-regulation of hepatic LDL receptor, consequently leading to an enhanced clearance of blood cholesterol (18).

The administration of HEFS and EEFS to OVX rats decreased serum cholesterol and TG compared with ovariectomized control rats. These findings were consistent with those of Sharma et al. (19), where fenugreek seed led to a significant decrease of serum cholesterol and TG. Hypolipidemic effect of fenugreek seed was partly related to the decrease in fat accumulation, up-regulation of LDL

receptor and decrease in insulin, which could decrease the activity of pyruvate dehydrogenase and acetyl-CoA carboxylase involved in lipogenesis as well as activity of 3-hydroxy-3-methylglutaryl-CoA reductase involved in cholesterol synthesis (20, 21). The role of fenugreek seed in the management of dyslipidemia and decreased risk of cardiovascular disease during the post menopause period was proposed (22). As compared with normal control rats, ovariectomy caused a considerable increase in the serum level of glucose and insulin. In the study of Mattace Raso et al. (23), no significant differences in serum glucose and insulin between OVX and normal rats were found. Shinmura et al. reported no statistical difference in serum glucose between OVX and normal rats (24). In contrast to the present study, the study of Ahmadi et al. (25) showed significantly decreased serum insulin in OVX rats as compared with control animals. In line with the present study, Posa et al. demonstrated that serum insulin significantly increased in OVX rats (26), which could be regarded as a marker for developing an insulin-resistant state. In addition, an increase in serum insulin may be due to the hyperactivity of the pancreatic islet cells to serum glucose (27). Serum glucose and insulin were considerably lower in OVX rats treated with HEFS and EEFS compared with OVX control animals. Zahedi et al. (28) revealed that plasma glucose in diabetic rats treated with fenugreek seed decreased compared to levels before treatment. It has been suggested that fenugreek seed causes a hypoglycemic effect through several mechanisms such as elevated glucose-stimulated insulin secretion by pancreatic beta cells (29) and activation of insulin receptor by tyrosine phosphorylation (30). It is likely that supplementary therapy of fenugreek seed extract could be beneficial for the improvement of glycemic control and insulin resistance after menopause.

In the present study, there was a significant decrease in the serum level of glucose and insulin in OVX rats exposed to estradiol when compared to OVX control rats. However, Ahmadi and Oryan showed that estradiol was an enhancer of serum insulin (25). You et al. identified that estradiol in OVX rats remarkably reduced serum glucose and insulin after six weeks (31). In addition, Li et al. showed that estrogen therapy might decrease blood insulin and glucose in menopausal females (32). It appears that estrogen by modulating insulin sensitivity or reducing insulin resistance (33) and lowering lipids (34) could be protective against metabolic abnormalities such as coronary artery disease (CAD) in menopausal females (31).

Ovariectomy led to a remarkable increase in serum Apelin in comparison to normal conditions. Boucher et al. showed that serum level of Apelin was considerably higher in moderately obese humans compared with the control group (6). Increased glucose in OVX rats may cause hyperactivity of pancreatic beta cells and increased insulin (27) and consequently increased Apelin (6). On the other hand, it has been suggested that Apelin has a role in decreasing the amount of glucose via enhanced

glucose utilization in muscles (35), and in inhibition of insulin secretion through induction of phosphatidylinositol 3-kinase and phosphodiesterase 3B (36). Therefore, increased serum Apelin in OVX animals appears to act partially as a regulatory mechanism; nevertheless, other possible theories need to be evaluated.

Serum level of Apelin was similar in OVX rats exposed to HEFS or EEFS and OVX control rats. Regarding the potential effects of fenugreek seed on lipids, glucose, and insulin (22, 37, 38), it appears that these effects may not be mediated by serum Apelin. However, more relevant investigations are required to verify this theory.

In OVX animals, serum Apelin remarkably decreased during estradiol treatment compared with the control group. Considering the fact that estradiol has a lowering effect on serum insulin (31, 32) and insulin can elevate Apelin production (6), it appears that reduced serum Apelin under estradiol treatment in OVX rats can be, to some extent, due to decreased insulin. Because of the mitogenic role of Apelin in the epithelial (39) and endothelial cells (40), this effect might be considered in agreement with the beneficial effects of hormone therapy in menopausal females (32). Nevertheless, the complicated role of Apelin during menopause needs to be further explored.

Overall, it appears that fenugreek seed might be effective against hyperglycemia, hyperlipidemia and insulin resistance in OVX rats without impact on serum Apelin. The 17- β estradiol could have similar effects along with possible inhibitory effects on serum Apelin. Since Apelin may affect metabolism via several different mechanisms, more related investigations are needed to determine the potential effect of Apelin on menopausal conditions.

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Authors' Contributions

1- Study concept and design: Mahmood Abedinzade and Korosh Khanaki. 2- Acquisition of data: Sima Nasri. 3- Analysis and interpretation of data: Masome Jamal Omid. 4- Drafting of the manuscript: Korosh Khanaki. 5- Critical revision of the manuscript for important intellectual content: Korosh Khanaki. 6- Statistical analysis: Bizhan Porramezan. 7- Administrative, technical and material support: Mahmood Abedinzade. 8- Study supervision: Mahmood Abedinzade.

References

1. Ouzounian S, Christin-Maitre S. [What is menopause?]. *Rev Prat.* 2005;55(4):363-8.
2. Topcuoglu A, Uzun H, Aydin S, Kahraman N, Vehid S, Zeybek G, et al. The effect of hormone replacement therapy on oxidized low density lipoprotein levels and paraoxonase activity in post-menopausal women. *Tohoku J Exp Med.* 2005;205(1):79-86.

3. Jouyandeh Z, Nayeibzadeh F, Qorbani M, Asadi M. Metabolic syndrome and menopause. *J Diabetes Metab Disord*. 2013;**12**(1):4.
4. Harlow SD, Gass M, Hall JE, Lobo R, Maki P, Rebar RW, et al. Executive summary of the Stages of Reproductive Aging Workshop + 10: addressing the unfinished agenda of staging reproductive aging. *Menopause*. 2012;**19**(4):387-95.
5. Sabia S, Fournier A, Mesrine S, Boutron-Ruault M-C, Clavel-Chapelon F. Risk factors for onset of menopausal symptoms: results from a large cohort study. *Maturitas*. 2008;**60**(2):108-21.
6. Boucher J, Masri B, Daviaud D, Gesta S, Guigné C, Mazzucotelli A, et al. Apelin, a newly identified adipokine up-regulated by insulin and obesity. *Endocrinology*. 2005;**146**(4):1764-71.
7. Ringström C, Nitter MD, Bennet H, Fex M, Valet P, Rehfeldt JF, et al. Apelin is a novel islet peptide. *Regul Pept*. 2010;**162**(1-3):44-51.
8. Yue P, Jin H, Xu S, Aillaud M, Deng AC, Azuma J, et al. Apelin decreases lipolysis via Gq, Gi, and AMPK-dependent mechanisms. *Endocrinology*. 2011;**152**(1):59-68.
9. Habchi M, Duvaillard L, Cottet V, Brindisi MC, Bouillet B, Beacco M, et al. Circulating Apelinin is increased in patients with type 1 or type 2 diabetes and is associated with better glycaemic control. *Clin Endocrinol (Oxf)*. 2014;**81**(5):696-701.
10. Cantley J. The control of insulin secretion by adipokines: current evidence for adipocyte-beta cell endocrine signalling in metabolic homeostasis. *Mamm Genome*. 2014;**25**(9-10):442-54.
11. Abedinzade M, Nikokar I, Nasri S, Nursabaghi F. Effect of Hexanic and Alcoholic Extracts of Fenugreek Seed in Male Diabetic Rats. *Zahedan J Res Med Sci*. 2013;**15**(6):50-3.
12. Handa T, Yamaguchi K, Sono Y, Yazawa K. Effects of fenugreek seed extract in obese mice fed a high-fat diet. *Biosci Biotechnol Biochem*. 2005;**69**(6):1186-8.
13. Kannappan S, Anuradha C. Insulin sensitizing actions of fenugreek seed polyphenols, quercetin & metformin in a rat model. *Indian J Med Res*. 2009;**129**(4):401-8.
14. Abedinzade M, Nasri S, Jamal Omodi M, Ghasemi E, Ghorbani A. Efficacy of Trigonella foenum-graecum Seed Extract in Reducing Metabolic and Inflammatory Alterations Associated With Menopause. *Iran Red Crescent Med J*. 2015;**17**(5).
15. Goss PE, Qi S, Cheung AM, Hu H, Mendes M, Pritzker KP. Effects of the steroidal aromatase inhibitor exemestane and the nonsteroidal aromatase inhibitor letrozole on bone and lipid metabolism in ovariectomized rats. *Clin Cancer Res*. 2004;**10**(17):5717-23.
16. Heidarpour M, Moghadam Jafari A, Kazemi Mehrjerdi H. Effects of pomegranate seed oil on oxidative stress parameters and lipid profiles in ovariectomized rats. *Iran J Vet Surg*. 2013;**8**(2):17-22.
17. Colditz GA, Willett WC, Stampfer MJ, Rosner B, Speizer FE, Hennekens CH. Menopause and the risk of coronary heart disease in women. *N Engl J Med*. 1987;**316**(18):1105-10.
18. Owen AJ, Roach PD, Abbey M. Regulation of low-density lipoprotein receptor activity by estrogens and phytoestrogens in a HepG2 cell model. *Ann Nutr Metab*. 2004;**48**(4):269-75.
19. Sharma MS, Choudhary PR. Hypolipidemic effect of fenugreek seeds and its comparison with atorvastatin on experimentally induced hyperlipidemia. *J Coll Physicians Surg Pak*. 2014;**24**(8):539-42.
20. Harris IR, Hoppner H, Siefken W, Farrell AM, Wittern KP. Regulation of HMG-CoA synthase and HMG-CoA reductase by insulin and epidermal growth factor in HaCaT keratinocytes. *J Invest Dermatol*. 2000;**114**(1):83-7.
21. Brownsey RW, Boone AN, Elliott JE, Kulpa JE, Lee WM. Regulation of acetyl-CoA carboxylase. *Biochem Soc Trans*. 2006;**34**(Pt 2):223-7.
22. Vijayakumar MV, Pandey V, Mishra GC, Bhat MK. Hypolipidemic effect of fenugreek seeds is mediated through inhibition of fat accumulation and upregulation of LDL receptor. *Obesity (Silver Spring)*. 2010;**18**(4):667-74.
23. Mattace Raso G, Santoro A, Russo R, Simeoli R, Paciello O, Di Carlo C, et al. Palmitoylethanolamide prevents metabolic alterations and restores leptin sensitivity in ovariectomized rats. *Endocrinology*. 2014;**155**(4):1291-301.
24. Shinmura K, Nagai M, Tamaki K, Bolli R. Loss of ischaemic preconditioning in ovariectomized rat hearts: possible involvement of impaired protein kinase C epsilon phosphorylation. *Cardiovasc Res*. 2008;**79**(3):387-94.
25. Ahmadi R, Oryan S. Effects of ovariectomy and estradiol valerate or progesterone on serum insulin level in rats. *Int J Med Med Sci*. 2013;**1**(6):263-6.
26. Pósa A, Szabó R, Kupai K, Csonka A, Szalai Z, Veszelka M, et al. Exercise training and calorie restriction influence the metabolic parameters in ovariectomized female rats. *Oxid Med Cell Longev*. 2015;**2015**:787063.
27. Liang K, Du W, Zhu W, Liu S, Cui Y, Sun H, et al. Contribution of different mechanisms to pancreatic beta-cell hyper-secretion in non-obese diabetic (NOD) mice during pre-diabetes. *J Biol Chem*. 2011;**286**(45):39537-45.
28. Zahedi Asl S, Farahnaz S, Ghasemi A, Zaree B. The effect of carbon tetrachloride extract of Trigonella foenum graecum seeds on glycogen content of liver in streptozotocin-induced diabetic rats. *Int J Endocrinol Metab*. 2007;**5**(2):70-5.
29. Broca C, Manteghetti M, Gross R, Baissac Y, Jacob M, Petit P, et al. 4-hydroxyisoleucine: effects of synthetic and natural analogues on insulin secretion. *Eur J Pharmacol*. 2000;**390**:345.
30. Vijayakumar MV, Singh S, Chhipa RR, Bhat MK. The hypoglycaemic activity of fenugreek seed extract is mediated through the stimulation of an insulin signalling pathway. *Br J Pharmacol*. 2005;**146**(1):41-8.
31. You MK, Rhuy J, Jeong KS, Bang MA, Kim MS, Kim HA. Effect of St. John's Wort (*Hypericum perforatum*) on obesity, lipid metabolism and uterine epithelial proliferation in ovariectomized rats. *Nutr Res Pract*. 2014;**8**(3):292-6.
32. Li C, Samsioe G, Borgfeldt C, Bendahl P, Wilawan K, Åberg A. Low-dose hormone therapy and carbohydrate metabolism. *Fertil Steril*. 2003;**79**(3):550-5.
33. Saglam K, Polat Z, Yilmaz M, Gulec M, Akinci SB. Effects of postmenopausal hormone replacement therapy on insulin resistance. *Endocrinology*. 2002;**18**(3):211-4.
34. Grodstein F, Stampfer MJ, Manson JE, Colditz GA, Willett WC, Rosner B, et al. Postmenopausal estrogen and progestin use and the risk of cardiovascular disease. *N Engl J Med*. 1996;**335**(7):453-61.
35. Dray C, Knauf C, Daviaud D, Waget A, Boucher J, Buleon M, et al. Apelin stimulates glucose utilization in normal and obese insulin-resistant mice. *Cell Metab*. 2008;**8**(5):437-45.
36. Guo L, Li Q, Wang W, Yu P, Pan H, Li P, et al. Apelin inhibits insulin secretion in pancreatic beta-cells by activation of PI3-kinase-phosphodiesterase 3B. *Endocr Res*. 2009;**34**(4):142-54.
37. Hannan JM, Ali L, Rokeya B, Khaleque J, Akhter M, Flatt PR, et al. Soluble dietary fibre fraction of Trigonella foenum-graecum (fenugreek) seed improves glucose homeostasis in animal models of type 1 and type 2 diabetes by delaying carbohydrate digestion and absorption, and enhancing insulin action. *Br J Nutr*. 2007;**97**(3):514-21.
38. Raghuram T, Sharma R, Sivakumar B, Sahay B. Effect of fenugreek seeds on intravenous glucose disposition in non-insulin dependent diabetic patients. *Phytother Res*. 1994;**8**(2):83-6.
39. Wang G, Anini Y, Wei W, Qi X, O. Carroll AM, Mochizuki T, et al. Apelin, a new enteric peptide: localization in the gastrointestinal tract, ontogeny, and stimulation of gastric cell proliferation and of cholecystokinin secretion. *Endocrinology*. 2004;**145**(3):1342-8.
40. Masri B, Morin N, Cornu M, Knibiehler B, Audigier Y. Apelin (65-77) activates p70 S6 kinase and is mitogenic for umbilical endothelial cells. *FASEB J*. 2004;**18**(15):1909-11.